The role of oval cells in chronic hepatitis type B and type C. A clinicopathologic study

A.C. Tsamandas¹, E.S. Felekouras², Stavroula Salakou³, Th. Kourelis¹, K. Tepetes², D.S. Bonikos¹, E. Papalampros², G.K. Michalopoulos³

SUMMARY

Aim: Oval hepatocytes, (OC) as a part of liver stem cells, are involved in the progress of liver disease and hepatocellular carcinoma (HCC) development, in experimental models. This study investigates the presence and possible role of OC in chronic hepatitis (CH) type B and C in humans.

Design: The study comprised 50 needle liver biopsies with CH and 50 wedge/needle normal liver biopsies as controls. CH cases included 29 HBV (all HBSAg+-14HbcAg+), and 21 HCV (PCR+) instances. HAI ranged from 3/18-15/18, and the architectural stage from 1 to 6 (6HBV/8HCV). Epidemiologic and laboratory data were obtained from medical records. The streptavidin-biotine technique was employed on paraffin sections using antibodies for cytokeratin 19 (CK19), glutathione-S-transferase- π (GST- π) and leukocyte common antigen (LCA). Cells with microscopic features of OC that were CK19 (+), or GST- π (+) and LCA (-) were scored. Oval cell percentage was expressed following morphometric analysis, and the results were correlated with clinical parameters, grade and stage of CH.

Results: Oval cells were present in all 50 liver biopsies with CH but in 0/50 controls. They were located in periportal

Department of Pathology¹ University of Patras School of Medicine, Patras Greece, First Surgical Clinic² University of Athens, Athens Greece, Department of Pathology³ University of Pittsburgh School of Medicine, Pittsburgh PA, USA

Author for correspondence:

Athanassios C. Tsamandas, MD, Department of Pathology University of Patras School of Medicine, 88 Agiou Demetriou str., 262 25 Patras, Greece, Tel: +30-61-271238, Fax: +30-61-994202, e-mail: tsamanda@otenet.gr areas; occasionally formation of small ducts was recorded. Oval cells were also recognized more often in association with fibrous bands and accompanied inflammatory infiltrates. The CK19+ cells varied from 20-90% and GST- π + cells from 20-75%, of the total oval cells numbers. The percentage of CK19+ and GST- π + cells were directly correlated with i) categories A, B, and D of Ishak score (p<0.01), ii) total Ishak score (p<0.01), iii) architectural grade (p<0.01), iv) transaminases values (p<0.05). Direct correlation was also seen between CK19 (+) and GST- π (+) cells (p<0.05).

Conclusions: Presence of oval hepatocytes is a common finding in liver biopsies from patients with CH type B or C. The significant association between oval cell percentage and severity of disease (HAI grade and stage) implies that OC proliferation is related to progress of liver disease and/or increased risk for hepatocellular carcinoma development.

Key words: oval cell, chronic hepatitis, carcinogenesis, fibrosis

INTRODUCTION

Hepatocellular carcinoma (HCC) is one of the most common human malignancies and its risk of development is significantly increased in chronic liver disease, especially in instances of chronic hepatitis type C and when cirrhosis is present.¹⁻⁵ After cirrhosis is established, liver regeneration following hepatocyte necrosis, is mediated by mature liver cell proliferation and alternatively by stem cell proliferation and differentiation into hepatocytes.⁶⁹

The origin of hepatic stem cell that produces progeny called oval cell in the adult liver of humans and animals, has been the subject of extensive research over the past several decades. Activation of the hepatic stem cell compartment is histologically detectable only when hepatocyte proliferation is suppressed.^{10,11} Oval cells are seen to arise in the periportal region of the liver and morphologically are small in size (approximately 10µm) and have a large nucleus-to-cytoplasmic ratio, with an oval shaped nucleus.¹² They have similarities to bile duct cells and hepatocytes with distinct isoenzyme profiles, expressing high levels of α -fetoprotein (AFP), certain keratin markers (cytokeratin 19-CK19), and γ -glutamyl transpeptidase.^{18,13-16} Immunohistochemical markers include antibodies such as OV6, CK19, GST- π , MK-2, and AFP.¹⁷⁻²¹ High levels of certain mRNAs like AFP and stem cell factor can also be expressed by oval cells.²⁰

Several previous studies have documented the emergence of oval cells following chemical injury, such as with ethionine and with galactosamine.7,22-24 Studies in rodent models of hepatocarcinogenesis suggest that activation of oval cells could be the mechanism by which the liver may replace destroyed parenchyma in chronic liver disease.^{6-9,25,26} In addition, experiments in animals exposed to a variety of carcinogenic regimens showed a uniform pattern of preneoplastic changes ultimately giving rise to HCC. In humans, oval cells have been reported in hepatitis B-associated HCC and chronic liver disease that associated with ductular proliferation.^{19,27,28} We performed the study mentioned in this report to detect the presen-ce of oval cells in cases of chronic hepatitis B or C severity and determine a possible relationship between the severity of liver disease and the number of oval cells.

MATERIALS AND METHODS

Patients

The study comprised 50 patients with chronic hepatitis (CH) proven by liver biopsy and diagnosed according to standard criteria.²⁹ They were 29 HBV (all HBSAg+-14HbcAg+), and 21 HCV (PCR+) instances. For the purposes of the study, all the epidemiological, clinical, and biochemical features pertaining these patients at the time of the diagnosis, were obtained by review of the clinical records, and are listed in table 1.

Methods

Conventional pathologic design

All liver tissues obtained by needle biopsy (14G) were fixed in 10% neutral formalin, embedded in paraffin and examined in multiple levels (10 sections of 4 μ m each). Histological study included hematoxylin and eosin (H&E), periodic acid Schiff (PAS) for bile duct evaluation), Masson's trichrome and reticulin. Several histoTable 1. Clinical and histologic characteristics of patients(N=100)

Sex	HBV 19M; 10F	HCV 29 21 16M; 5F
Mean (range)	52 (30-70)	49 (25-60)
HAI score, mean (range)		
Category A	3 (1-4)	2 (1-4)
Category B	1 (0-3)	1 (0-3)
Category C	2 (1-4)	2 (1-4)
Category D	3 (1-4)	3 (1-4)
Total score	9 (3-15)	8 (3-15)
Stage		
1	8	3
2	4	2
3	4	2
4	4	3
5	3	3
6	6	8
Laboratory values mean (ra	nge)	
AST	163 (98-205)	144 (91-198)
ALT	177 (90-210)	162 (89-201)

CH: chronic hepatitis, HBV: hepatitis B, HCV: hepatitis C, HAI: hepatitis activity index

logic features were assessed in each biopsy in a blind fashion, and finally the Hepatitis Activity Index (HAI) was applied and the architectural grade was recorded in each instance.³⁰ For control purposes 50 wedge or needle biopsies from normal liver parenchyma were used. These were obtained from 50 patients during operation for cholelithiasis, without co-existing choledocholithiasis.

Immunohistochemistry

All 100 liver biopsies (50 with CH and 50 controls) included in the study were stained for presence or absence of GST- π , CK19 and LCA antigens, using a standard streptavidin-biotin immunohistochemical method. For CK19 expression microwave antigen retrieval was used. Primary antibodies used included: monoclonal mouse antibody to CK19 (Novocastra UK), in a dilution 1:80, polyclonal antibody GST- π (Novocastra, UK), in a dilution of 1:80 and monoclonal antibody to LCA (Biogenex, UK) ready for use.

Measurement of oval cell proliferation

Each liver biopsy was scanned in a low power and

cells that fullfilled the morphologic criteria of oval cells and displayed cytoplasmic positivity to CK19 or GST- π but were negative for LCA, were scored. Cell counts were performed manually at a X400 magnification using a 10x10-microscope grid. Both the numbers of immunoreactive cells and the total number of oval cells (at least 500 cells) were determined by visual inspection of five non-overlapping different fields per section. The percent of immunoreactive (CK19+ or GST- π + cells) was based on the ratio of positive cells to the total number of cells counted. The variance in oval cell counts from section to section in the same biopsy was <10%. The average of these scores was then taken.

Statistical analysis

Results were expressed as the mean \pm and standard deviation. The Mann Whitney test was employed to determine whether the differences between groups were significant. A p-value < 0.05 was set as significant.

RESULTS

Liver pathology

All 50 needle liver biopsies included in the study, contained more than 4 portal tracts and considered sufficient for pathologic evaluation. Table 1 demonstrates the value of each HAI parameter, total HAI score and HAI stage for chronic hepatitis instances. Total HAI score ranged from 3/18 (3HBV and 2HCV cases) to 15/18 (4HBV and 2HCV cases). Evaluation of Masson's trichrome and reticulin stain revealed HAI stage 6 in 6HBV and 8HCV cases.

Immunohistochemical results

Oval cells (CK19+ or GST- π + and simultaneously LCA negative cells) were present in all 50 biopsies with chronic hepatitis but in none of the control biopsies. They were located in periportal areas and occasionally formed ductular structures (Fig 1, 2). Oval cells were found in close proximity with fibrous septa, surrounding regenerative nodules, or associated with inflammatory cell infiltrates (Fig 3). GST- π was also expressed in mature hepatocytes (Fig 4). The number of CK19 positive oval varied from 20 ± 2 to 90 ± 8 in cases of chronic hepatitis type B and from 22 ± 4 to 89 ± 6 in chronic hepatitis type B and from 21 ± 4 to 75 ± 3 in chronic hepatitis type C instances.

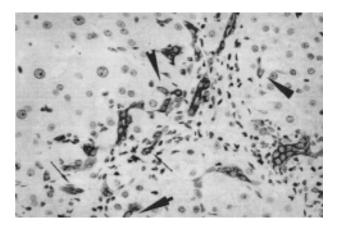


Figure 1. Photomicrograph of immunohistochemistry for CK19+ oval cells (thick arrows) that form ductular structures (thin arrows) in a case of chronic hepatitis type B (ABC x400).

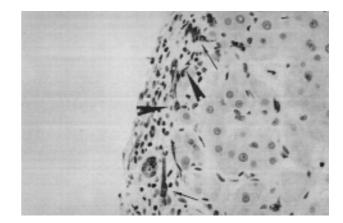


Figure 2. Photomicrograph of immunohistochemistry for GST- π + oval cells (thick arrows) that form ductular structures (thin arrows) in a case of chronic hepatitis type B (ABC x 400).

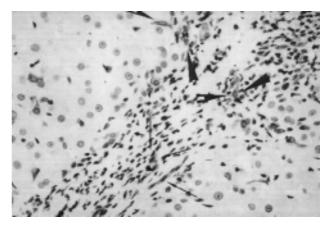


Figure 3. Photomicrograph of immunohistochemistry for CK19+ oval cells (thick arrows) present within fibrous septa, that accompany lymphocytic inflammation in a case of chronic hepatitis type C. Note that oval cells form ductular structures (thin arrows). (ABC x 200).

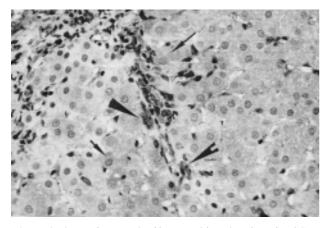


Figure 4. Photomicrograph of immunohistochemistry for GST- π - expressed within oval cells (thick arrows) and hepatocytes (thin arrows) (ABC x 200).

Correlation between parameters

Positive correlations were observed between CK19+ oval cells and i) categories A, B, and D of HAI score (p<0.01 in each instance), ii) total HAI score (p<0.01), iii) architectural grade (p<0.01), iv) transaminases values (p<0.05 in each instance). Similar correlations observed between GST- π + cells with each parameter of HAI score, total HAI score, hepatitis grade and transminases values. In addition the numbers of CK19+ cells were positively correlated with GST- π + cells (p<0.05). No significant correlation (p>0.05) was recorded between CK19+ and GST- π + cells and cause of hepatitis when the values were matched for each parameter of HAI, total HAI or hepatitis grade.

DISCUSSION

This study demonstrates that oval cells are frequently present in patients with chronic hepatitis B or C. In addition, oval cell numbers increase significantly with progression of disease severity (grade and stage) in both groups studied, suggesting that oval cell proliferation is not disease-specific but occurs in response to progressive liver injury and fibrosis. This concurs with the hypothesis that oval cell activation is associated with increased risk for development of HCC with advancing liver disease, particularly when cirrhosis is present.¹⁹

The nature, site, and cell type origin of the "oval cells" that emerge in the liver after different types of injury has been a subject of several studies.^{8,10,12-18,20} The appearance of both bi-liary and hepatocytic markers in these cells has reinforced the concept of the hepatic stem cell, and many investigators in the field consider these oval cells

as the immediate progeny of a putative hepatic stem cell.18 Chemical injury to the liver induced by a variety of agents such as galactosamine and ethionine cause an increase in the numbers of oval cells.^{7,22,24}

In animal models oval cell proliferation is associated with the presence of inflammatory cells within the liver.^{26,31} When oval cells are isolated from animals placed on a choline-deficient, ethionine-supplemented diet, they rapidly deteriorate in culture suggesting that exogenous factors are required for their survival and proliferation.²⁶ It has been referred that oval cells presence is reduced in cases of experimental selective periportal damage even when mature hepatocyte proliferation has been inhibited.11,24 In this and other previous studies, oval cells were often found in close association with fibrous tissue in the liver, often proliferating along the tracts from the expanded portal regions, and along the limiting plates surrounding regenerative nodules.¹⁹ In addition, oval cells were often observed in close association with inflammatory cells, in patients with chronic hepatitis B or C. These observations suggest that cytokines or other factors produced by inflammatory cells or cells associated with the development of fibrosis, such as liver myofibroblasts and Kupffer cells, may be required to stimulate oval cell proliferation and migration.

Oval cell detection in human liver diseases is based on the presence of cells with the typical histopathological appearance combined with appropriate immunohistochemical markers. Several immunohistochemical markers have been described for oval cells including OV6, CK19, GST-π, MK-2, and AFP.^{17-21,32} The combined use of immunohistochemistry and morphology to characterize oval cells, and the exclusion of inflammatory cells (LCA negative cells) in serial sections) lead to reliable results regarding oval cells presence. This study shows that GST- π and CK19 are reliable markers for oval cells. The numbers of CK19+ oval cells were higher when compared with GST- π + cells (when the cases were matched for other histopathologic features), and this may reflect the heterogeneity of the oval cell population in terms of their developmental maturity or their commitment to either the hepatocytic or biliary lineage.

The significance of the ductule-like structures formed by oval cells in various disease states has not yet been clarified. Previous studies in rodents have demonstrated that oval cells can proliferate and form ductular-like structures during carcinogenesis and biliary obstruction.^{25,26,32} Some of the cells in the ductule-like structures also express the adult L-pyrouvate kinase isoenzyme (L-PK), with a small population coexpressing M2-PK and L-PK.^{25,26,32} These observations suggest that some cells within the ductular structures may be capable of progressing along the hepatocyte lineage. Formation of ductal structures has been observed during liver regeneration following selective periportal necrosis after pretreatment with inhibitors of mature hepatocyte proliferation.^{11,24} Possible explanations could be that a) the forerunners of the oval cells cannot "colonize" the immediate periportal site and lead to the oval cell response due to the damage of the overall tissue environment and b) that the canals of Herring (considered by most investigators as the compartment of biliary epithelium most likely to be associated with the oval cell response) are also damaged as "innocent bystanders" in chronic hepatitis cases as the disease progresses.

In conclusion, this study demonstrates that presence of oval cells is a common finding in liver biopsies from patients with chronic hepatitis type B or type C. The significant association between oval cell percentage and severity of disease (HAI grade and stage) implies that proliferation of those cells is related to progress of liver disease and/or increased risk for hepatocellular carcinoma development.

REFERENCES

- Kew MC. Hepatic tumors and cysts. Sleisinger and Fordtran's gastrointestinal and liver diseases: pathophysiology/diagnosis/management. Edited by M Feldman, BF Scharschmidt, MH Sleisinge, Philadelphia, WB Saunders, 1998, pp1364-1387.
- MacSween RNM, Scott AR. Hepatic cirrhosis: a clinicopathological review of 520 cases. J Clin Pathol 1973; 26:936-942.
- Colombo M, Kuo G, Choo QL, Donato MF, Del Ninno E, Tommasini MA, Dioguardi N. Prevalence of antibodies to hepatitis C virus in Italian patients with hepatocellular carcinoma. Lancet 1989; 2:1006-1008.
- Roudot-Thoraval F, Bastie A, Pawlotsky JM, Dhumeaux D. Epidemiological factors affecting the severity of hepatitis C virus-related liver disease: a French survey of 6,664 patients. The Study Group for the Prevalence and the Epidemiology of Hepatitis C virus. Hepatology 1997; 26:485-490.
- Tsukuma H, Hiyama T, Tanaka S, Nakao M, Ybuuchi T, Kitamura T, nakanishi K, Fujimoto I, Inoue A, Yamazaki H, Kawashima T. Risk factors for hepatocellular carcinoma among patients with chronic liver disease. N Engl J Med 1993; 328:1797-1801.
- Sell S, Dunsford HA. Evidence for the stem cell origin of hepatocellular carcinoma and cholangiocarcinoma. Am J Pathol 1989; 134:1347-1363.
- 7. Lemire JM, Shiojiri N, Fausto N. Oval cell proliferation and the origin of small hepatocytes in liver injury induc-

ed by D-galactosamine. Am J Pathol1991; 139:535-552.

- Evarts RP, Nagy P, Marsden E, Thorgeirsson SS. A precursor-product relationship exists between oval cells and hepatocytes in rat liver. Carcinogenesis 1987; 8:1737-1740.
- Hixson DC, Fowler LC. Development and phenotypic heterogeneity of intrahepatic biliary epithelial cells. Biliary and pancreatic ductal epithelia: pathobiology and pathophysiology. Edited by AE Sirica, DS Longnecker, New York, Marcel Dekker, 1997, pp1-40.
- Evarts RP, Nagy P, Nakatsukasa H, Marsden E, Thorgeirsson SS. In vivo differentiation of rat oval cell hepatocytes. Cancer Res. 1989; 49:1541-1547.
- Petersen BE, Zajac VF, Michalopoulos GK. Hepatic oval cell activation in response to injury following chemically induced periportal or pericentral damage in rats. Hepatology 1998; 27(4): 1030-1038.
- Farber E. Similarities in the sequence of early histological changes induced in the liver of the rat by ethionine, 2acetylaminofluorene, and 3'-methyl-4-dimethylaminoazobenzene. Cancer Res. 1956; 16:142-148.
- Germain L, Noel M, Gourdeau H, Marceau N. Promotion of growth and differentiation of rat ductular oval cells in primary culture. Cancer Res. 1988; 48:368-378.
- Shiojiri N, Lemire JM, Fausto N. Cell lineage and oval cell progenitors in rat liver development. Cancer Res. 1991; 51:2611-2620.
- Dabeva MD, Shafritz DA. Activation, proliferation and differentiation of progenitor cells into hepatocytes in the D-galactosamine model of the liver regeneration. Am J Pathol 1993; 143:1606-1620.
- Fausto N. Liver stem cells. In Arias IM, Boyer JL, Fausto N, Jacoby WB, Schachter DA, Shafritz DA Eds. The Liver: Biology and Pathobiology, 3rd ed. New York: Raven, 1994:1501-1518.
- Faris RA, Monfils BA, Dunsford HA, Hixson DC. Antigenic relationship between oval cells and subpopulation of hepatic foci, nodules, and carcinomas induced by the "resistant hepatocyte" model system. Cancer Res. 1991; 51:1308-1317.
- Thorgeirsson SS, Evarts RP, Bisgaard HC, Fujio K, Hu Z. Hepatic stem cell compartment: activation and lineage commitment. Proceedings of the society for experimental biology and medicine 1993; 204:253-260.
- Lowes KN, Brennan BA, Yeoh GC, Olynyk JK. Oval cells numbers in human chronic liver diseases are directly related to disease severity. Am J Pathol 1999;154:537-541.
- Fujio K, Evarts RP, Hu Z, Marsden ER, Thorgeirsson SS. Expression of stem cell factor and its receptor, c-kit, during liver regeneration from putative stem cells in the adult rat. Lab Invest 1994; 704:511-516.
- Vandersteenhoven AM, Burchette J, Michalopoulos GK. Characterization of ductular hepatocytes in end-stage cirrhosis. Arch Pathol Lab Med 1990; 114:403-406.
- 22. Shinozuka H, Lombardi B, Sell S, Iammario RM. Earlynhistological and functional alterations of ethionine liver carcinogenesis in rats fed a choline-deficient diet. Cancer Res 1978; 38:1092-1098.
- 23. Novikoff P, Yam A, Oikawa I. Blast-like cell compartme-

nt in carcinogen-induced prolferating bile ductules. Am J Pathol 1996; 148:1473-1492.

- 24. Felekouras E, Tsamandas A, Tepetes K, Sigala F, Salakou S, Papalampros E, Michalopoulos G. Oval cell activation in respons to injury following different models of hepatic damage with or without inhibition of mature hepatocyte proliferation. Modern Pathology; 13(1):183A, 2000.
- 25. Tian YW, Smith PG, Yeoh GC. The oval-shaped cell as a candidate for liver stem cell in embryonic, neonatal and precancerous liver: identification based on morphology and immunohistochemical staining for albumin and pyrouvate kinase isoenzyme expression. Histochem Cell Biol 1997;107:243-250.
- 26. Tee LB, Kirilak Y, Huang WH, Morgan RH, Yeoh GC. Differentiation of oval cells into duct-like cells in preneoplastic liver of rats placed on a choline-deficient diet supplemented with ethionine. Carcinogenesis 1994; 15:2747-2756.
- 27. Ray MB, Mendenhall CL, French SW, Gartside PS. Bile duct changes in alcoholic liver disease. The Veterans

Administration Cooperative Study Group. Liver 1993; 13:36-45.

- Hsia CC, Evarts RP, Nakatsukasa H, Marsden ER, Thorgeirsson SS. Occurrence of oval type cells in hepatitis B virus associated human hepatocarcinogenesis. Hepatology 1992; 16:1327-1333.
- 29. Lee RG. Chronic hepatitis, In Lee RG ed. Diagnostic Liver Pathology, 1st ed. St. Louis, Mosby-Year book 1994,57-79.
- 30. Ishak K, Baptista A, Bianchi L, et al. Histological grading and staging of chronic hepatitis. Journal of Hepatol 1995; 22(6):696-9.
- 31. Tee LB, Smith PG, Yeoh GC. Expression of α, μ and pi class glutathione S-transferases in oval and ductal cells in liver of rats placed on a choline-deficient, ethionine-supplemented diet. Carcinogenesis 1992; 13:1879-1885.
- 32. Bisgaard HC, Nagy P, Ton PT, Hu Z, Thorgeirsson SS. Modulation of keratin 14 and α-fetoprotein expression during hepatic oval cell proliferation and liver regeneration. J Cell Physiol 1994; 159:475-484.