# Protective effect of *Melissa officinalis* against acetic acid-induced ulcerative colitis in rat models: an experimental study

Reza Shahriarirad<sup>a,b</sup>, Amirhossein Erfani<sup>a,b</sup>, Fatemeh Nekouei<sup>a</sup>, Sarvin Seifbehzad<sup>a</sup>, Masood Hosseinzadeh<sup>d</sup>, Bahador Sarkari<sup>e</sup>, Nader Tanideh<sup>f</sup>, Omid Koohi-Hosseinabadi<sup>g</sup>, Nour Nassour<sup>c</sup>, Soheil Ashkani-Esfahani<sup>c</sup>

Shiraz University of Medical Sciences, Shiraz, Iran; Massachusetts General Hospital, Boston, MA, USA

# Abstract

**Background** Inflammation and oxidative activities within the gut play major roles in the pathogenesis of ulcerative colitis (UC). We aimed to determine the effect of *Melissa officinalis*, an antioxidant and anti-inflammatory agent, on the colon histological characteristics in acetic acid (AA)-induced UC in rat models.

**Methods** Thirty-six male rats with AA-induced colitis were divided into 5 groups: no treatment (AA); daily treatment with 300 mg/kg *Melissa officinalis* orally (MO) and rectally (MR); and 100 mg/kg mesalamine orally (AO) and rectally (AR). Macroscopic and histopathological evaluation of the colon, along with a biochemical laboratory evaluation, were performed 10 days after UC induction.

**Results** All treatment groups demonstrated lower macroscopic grading scores compared to the AA group. After treatment with MO, 42.9% of cases demonstrated no macroscopic changes, while 14.3% demonstrated only mucosal erythema. In the MR group 28.6% of rats had no changes in their mucosal lining and 28.6% had only mucosal erythema. Following histopathological evaluation, the AO group had lower scores regarding the severity of ulcer, inflammation, destruction, crypt abscess, and disorganization compared to the MO group. (P=0.02) The MR group demonstrated lower microscopic scores compared to the MO group, and also lower macroscopic scores compared to the AR group, although not significantly (P>0.05).

**Conclusions** Both oral and topical administration of *Melissa officinalis* have satisfactory healing properties compared to mesalamine, with topical route having better results. Therefore, further studies are needed to establish the benefit of *Melissa officinalis* administration (both orally and topically) within a UC treatment protocol.

Keywords Anti-inflammatory effects, *Melissa officinalis*, mesalamine, treatment, ulcerative colitis

Ann Gastroenterol 2023; 36 (XX): 1-7

Conflict of Interest: None

Received 15 March 2023; accepted 15 September 2023; published online 30 October 2023

Correspondence to: Reza Shahriarirad, MD, Thoracic and Vascular Surgery Research Center, Shiraz University of Medical Sciences, Shiraz, Iran, e-mail: R.shahriari1995@gmail.com

DOI: https://doi.org/10.20524/aog.2023.0836

This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 4.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms

# Introduction

Ulcerative colitis (UC) is a form of chronic inflammatory bowel disease (IBD) that disrupts the mucosal barrier of the gastrointestinal tract, and is also known to increase the risk of colorectal cancer [1]. Furthermore, there has been a report of increased incidence and prevalence of the disease [2]. The incidence also varies among countries, with the annual incidence of UC ranging from 8.8-23.1 per 100,000 personyears in North America, 7.3-17.4 in Oceania, and 0.6-24.3 per 100,000 person-years in Europe [3,4].

The exact cause of UC has not yet been clarified; however, studies have established certain risk factors, such as stress,

smoking, familial history and prior appendectomy, that could play a role in its development [2]. The current standardized treatment of UC includes the administration of corticosteroids (e.g., prednisolone), aminosalicylates (e.g., 5-amino salicylic acid [5-ASA], sulfasalazine, mesalamine), immunomodulator agents and antibiotics. However, those medications have been shown to have several bothersome side effects, including headache, nausea, abdominal pain, lung infection, inflammation of the pancreas, and renal damage [5]. As it is hypothesized that the body's immune system, specifically its inflammatory and oxidative responses, contribute to the development of UC, many have attempted to study substances and agents that act on those responses and thus might help in the treatment of the disease, while mitigating side-effects [6]. Many efforts have been made to find more effective and benign treatments, focused on reducing the inflammatory and oxidative reactions in the process of UC development [7,8]. It is assumed that natural products with proven anti-inflammatory and antioxidative effects might have the potential to act as an alternative treatment for UC, with fewer side-effects [9,10].

*Melissa officinalis* (Melissa), also known as lemon balm, is a wild herb that is known to have special effects, such as relaxation, relief of nervousness, reduction in dizziness and headache, provision of energy and facilitation of digestion. The infusion and topical lotion from Melissa leaves extract are effective in relieving pain and healing wounds and injuries [11]. Melissa has also shown anti-inflammatory, antioxidant and antibacterial effects that may also be useful in treating diseases that are based on inflammatory and oxidative responses of the body, such as UC [11,12]. In this study, we aimed to evaluate the effects of Melissa on the pathological parameters and inflammatory markers of acetic acid (AA)-induced UC in rats.

## **Materials and methods**

#### Plant extraction and drug preparation

Melisa leaves were supplied by the SIMR company, Shiraz, Iran (Voucher number: 31112) as a dried powder. The drugextract ratio was 60:1, the dried extract corresponding to 1.63% of the primary raw plant's leaf material. The hydroalcoholic extract (65% v/v) was obtained by macerating 20 g of the herbal material for 1 week in the solution at 40°C in darkness. The solution was

<sup>a</sup>Student Research Committee, Shiraz University of Medical Sciences, Shiraz, Iran (Reza Shahriarirad, Amirhossein Erfani, Fatemeh Nekouei, Sarvin Seifbehzad); <sup>b</sup>Thoracic and Vascular Surgery Research Center, Shiraz University of Medical Sciences, Shiraz, Iran (Reza Shahriarirad, Amirhossein Erfani); Foot & Ankle Research and Innovation Laboratory (FARIL), Massachusetts General Hospital, Boston, MA, USA (Nour Nassour, Soheil Ashkani-Esfahani); <sup>d</sup>Department of Pathology, Shiraz University of Medical Sciences, Shiraz, Iran (Masood Hosseinzadeh); <sup>e</sup>Department of Parasitology and Mycology, Shiraz University of Medical Sciences, Shiraz, Iran (Bahador Sarkari); <sup>(Stem</sup> Cell Technology Research Center, Shiraz University of Medical Sciences, Shiraz, Iran (Nader Tanideh); <sup>g</sup>Center of Comparative and Experimental Medicine, Shiraz University of Medical Sciences, Shiraz, Iran (Omid Koohi-Hosseinabadi) then decanted from the extracted leaf residues, filtered and stored at -4°C. The concentration was 100 mg/mL, with reference to the initial dried herbal material. The vehicle was prepared using carboxy-methylcellulose (CMC) 0.3% (v/w) solution, based on previous reports [13]. In accordance with a previously conducted study, the concentration of 300 mg/kg of Melissa extract was chosen for the main experiment, to be administered using a CMC vehicle both rectally and orally [14-16].

Before performing the main experiment, in a pilot study, oral and rectal administrations of the vehicle were evaluated and compared with an untreated group of rats with AAinduced colitis. No beneficial effect of the vehicle was seen in comparison with the rats that received no treatment.

## Study design

The sample size was calibrated based on previous studies, and while assessing the risk of drop-out [17,18]. The Ethics Committee of Shiraz University of Medical Sciences, Shiraz, Iran approved the experimental protocol, and all the criteria for taking care of laboratory animals outlined in the "Guide for the Care and Use of Laboratory Animals" were applied (Ethical code: IR.SUMS.REC.1394.s1101). All efforts were made to keep animal distress to a minimum and to use only the number of animals essential to attain reliable results.

The animals were maintained under standard conditions (12 h light/dark cycle;  $24\pm3^{\circ}$ C, 45-55% humidity) and free access to standard food and water *ad libitum*. They were acclimatized to laboratory conditions for a week prior to the experiment. The animal study was performed during the daylight portion between 09:00 and 12:00 am, to avoid possible circadian impacts. The health status and body weight of animals were monitored daily and a loss of more than 20% of body weight was considered the threshold for a humane endpoint (none of the subjects met this criterion).

# **Animal grouping**

Thirty-six male Wistar rats  $(180\pm20 \text{ g})$  were obtained from the animal house of Shiraz university of medical sciences, Shiraz, Iran. The animals were divided into 5 groups: the control group (AA; n=4) consisted of rats with AA-induced colitis, which received no treatment; experimental groups MO (n=8) and MR (n=8), in which colitis was induced and which received 300 mg/kg Melissa solution daily orally and rectally, respectively; and the AO (n=8) and AR groups (n=8), which received a dose of 100 mg/kg mesalamine (5-ASA) orally and rectally, respectively.

#### Intervention

In accordance with previous experiments, all the treatments were started 4 days before induction of colitis, and on day 5 of the study colitis was induced [19]. One of the standardized experimental models of UC is through the induction of colitis by AA [20]. The oral treatments were given for 10 days using oral gavage; the rectal administration was performed using a 2-mm diameter polypropylene tube inserted into the colon to a distance of 5-8 cm, up to the limit where resistance was detected.

For induction of colitis, on the fourth day of the study the animals were fasted overnight with access to water *ad libitum*. On the fifth day, after 2 hours' administration of the treatments, the rats were anesthetized by ether inhalation and a polypropylene tube (2-mm diameter) was inserted through the rectum of the animal into the colon to a distance of 6-8 cm, depending on the body length. An AA (Sigma Aldrich, St. Louis, USA) solution (2 mL, 3% v/v) in 0.9% normal saline was instilled into the colon and the animal was maintained in a supine Trendelenburg position for about 30 sec to prevent leakage of the solution [19]. This method of AA-induction of colitis has also been examined in previous investigations [20-22].

#### **Data collection**

The treatments were continued until day 10 of the experiment, when the animals were anesthetized with ether inhalation and blood was collected by cardiac puncture for biochemical evaluations, including C-reactive protein (CRP), superoxide dismutase (SOD), white blood cell count (WBC), hemoglobin (HGB), and platelets. The enzymatic activities of SOD were based on the method developed by Misra and Fridovich [23]. Subsequently, we sacrificed the animals by decapitation, and their colons was dissected. A longitudinal incision was made to remove and open the distal part of the colon, approximately 8 cm. The mucosa was cleaned with saline solution, and mucosal injury was evaluated (macroscopically) in accordance with a previously described method by Millar et al. This uses an arbitrary scale with a 0-4 range to assign inflammation ratings based on the clinical characteristics of the colon: 0, no macroscopic alterations; 1, just mucosal erythema; 2, mild mucosal edema, slight bleeding, or minor erosions; 3, moderate edema, slight bleeding ulcers, or erosions; and 4, severe ulceration, edema, and tissue necrosis [24]. Additionally, samples were kept in 10% formalin for histological analysis.

Colonic samples were collected 2-4 cm from the anus. The tissue was then fixed in phosphate-buffered formaldehyde, embedded in paraffin, processed into 5-mm sections, stained with hematoxylin and eosin, examined under a light microscope, and eventually graded by an expert pathologist in a blinded fashion. The degree of the inflammatory reaction in the tissue was assessed using a histological grading system. Depending on the severity of changes, each parameter assessed was scored from 0-3 (0, no change; 1, mild; 2, moderate; 3, severe). The factors taken into consideration and subjectively graded included ulceration, inflammatory cell infiltration, mucosa damage, disarray and crypt abscess. Fig. 1 shows 2 colon samples from the MR group and Fig. 2 shows a sample of AA-induced colitis in different groups of the study.



Figure 1 Macroscopic evaluation of the excised distal part of the colon with no macroscopic change (top), compared with severe ulceration, edema and tissue necrosis in 2 subjects treated with rectal administration of *Melissa officinalis* (MR group)

#### **Statistical analysis**

SPSS software (v. 26) was used for the analysis. The data were checked for normal distribution and reported as mean±standard deviation or median and interquartile range. Data were analyzed using Fisher's exact test for descriptive data and using an independent sample t-test, Mann-Whitney *U* for 2-parameter evaluation, and one-way analysis of variance (ANOVA) or Kruskal-Wallis tests for multiple parameter evaluations. A *post hoc* test was used for intergroup comparison. A P-value <0.05 was considered as statistically significant.

# Results

Out of a total of 36 rats, 7 died after the induction of AA: 1 in the MR group; 1 in the MO group; 2 in the AR group; and 3 in the AO group. There was no significant difference among all groups regarding their initial weight (P=0.71). Based on repeated measures ANOVA, all groups developed a significant decrease in weight after the administration of AA up to the eighth day (P<0.001) (Fig. 3); however, this change was not statistically significant (P=0.11).

The outcomes of the macroscopic evaluation are reported in Table 1. AO had significantly lower scores compared to all groups except the MO group (P=0.11). The MR group also demonstrated lower scores compared to the AR group, although the difference was not significant (P=0.87).

Based on microscopic evaluation, the AO group had significantly lower histopathological scores regarding the severity of ulcer, inflammation, crypt abscess, destruction and disorganization compared to the MO group. The AO group mainly demonstrated no or mild changes, while the MO group showed a level of mild to severe changes in each of the abovementioned parameters (Table 1) (P=0.02; P=0.01; P=0.004; P=0.01; P=0.01, respectively). Although the MR group demonstrated higher microscopic scores than the AR group in the histopathological evaluation of the abovementioned factors, this difference was not statistically significant (P=0.65; P=0.81; P=0.84; P=0.39; P=0.21, respectively).

Biochemical data revealed no significant differences either among all treated groups, or compared to the AA group. The

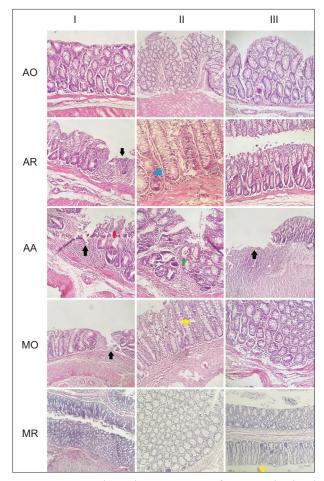


Figure 2 Hematoxylin and eosin staining of acetic acid-induced ulcerative colitis in the different groups. AO: orally administered mesalamine. I, II, III: no gland destruction no disorganization and no inflammation (normal mucosal and gland architecture). AR: rectally administered mesalamine. I: focal surface ulceration; II: mild active colitis (cryptitis); III: no destruction and gland disorganization. AA: without treatment. I: gland destruction and mucosal ulceration; II: crypt abscess and the overall slide demonstrates disorganization; III: surface ulceration. MO: orally administered Melissa officinalis (Melissa). I: ulceration and gland destruction; II: mucosal inflammation; III: mild disarray of gland architecture. MR: rectally administered Melissa. I: Mild disorganization of glands; II: no change in gland and mucosal architecture; III: mild mucosal inflammation. Black arrows show focal surface ulceration; yellow arrow shows mucosal inflammation; blue arrows show mild active colitis (cryptitis); green arrows show crypt abscess; red arrow shows gland destruction

outcomes of the biochemical laboratory examinations are shown in Table 1.

# Discussion

The administration of Melissa, a substance known for its anti-inflammatory and antioxidative effects, showed significant results in reversing the damage to the intestinal

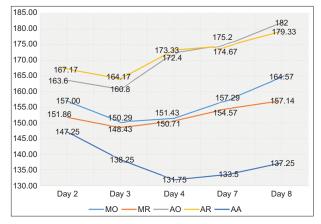


Figure 3 Comparison of the weight changes of rats with ulcerative colitis throughout the 5-day period of the study following treatment with *Melissa officinalis* and mesalamine in comparison to the control group (AA). The groups are as follows: *Melissa officinalis* Oral (MO), received 300 mg/kg oral treatment with *Melissa officinalis; Melissa officinalis*; rectal (MR), received 300 mg/kg rectal treatment with *Melissa officinalis;* 5-aminosalicylic acid (5-ASA) oral (AO) received 100 mg/kg oral treatment with mesalamine; 5-ASA rectal (AR) received 100 mg/kg rectal treatment with mesalamine; acetic acid (AA) denotes untreated animals with induced colitis

mucosa due to AA. In addition, 42.9% of cases in the MO group demonstrated no macroscopic changes, while 14.3% demonstrated only mucosal erythema. The MO group had no significant differences from the reference group (AO) regarding macroscopic features. On the other hand, the rectal administration of Melissa (MR) was superior to rectal mesalamine (AR group); although these differences were non-significant. The similarities in the healing properties of these medications cannot be overlooked. Furthermore, our results confirmed that oral administration of both Melissa and mesalamine was superior to their topical equivalents in regard to the resulting macroscopic changes. Overall, we advise the administration of Melissa (both orally and topically) as an addon or even an alternative treatment in UC.

In this study, the oral administration of mesalamine (AO group) demonstrated the most satisfactory results among all evaluated parameters, consistent with its administration as the reference drug in the management of UC [25]. Many studies have indicated that administering antioxidative and anti-inflammatory agents to the gut leads to improvements in the course of UC and reduces the disease's morbidity and mortality, in both experiments and human trials [19,26]. The application of herbal medicine in UC has been widely reported in the literature. Some of the administered treatments include Gegen Qinlian Decoction, fuzi-ganjiang, *Ramak* and *Cupressus sempervirens*, which are administered because of their antioxidant, immune boosting, anti-inflammatory and healing properties [27-31].

Although Melissa's therapeutic properties have been reported in the literature, studies supporting the administration of Melissa in the treatment of IBD are very limited [32,33]. Commercial capsules such as Melipass<sup>°</sup>, which is a flavonoid-rich phytotherapeutic agent based on

Factor	Group*					P-value**
	MO; n=7	MR; n=7	AO; n=5	AR; n=6	AA; n=4	
		Gross	examination			
Macroscopic Grade ***						
Total	1.0 [2]	1.0 [4]	0 [0]	1.5 [2]	2.5 [3]	0.04
0	3 (42.9)	2 (28.6)	5 (100)	0 (0)	0 (0)	0.07
1	1 (14.3)	2 (28.6)	0 (0)	3 (50.0)	1 (25.0)	
2	2 (28.6)	1 (14.3)	0 (0)	1 (16.7)	1 (25.0)	
3	0(0)	0 (0)	0 (0)	2 (33.3)	1 (25.0)	
4	1 (14.3)	2 (28.6)	0 (0)	0 (0)	1 (25.0)	
		Histo	ological grade			
Ulcer						
Total	1.0 [2]	1.0 [1]	0 [0]	0 [1]	0.5 [2]	0.16
0: No Change	2 (28.6)	3 (42.9)	5 (100)	4 (66.7)	2 (50.0)	0.67
1: Mild	2 (28.6)	3 (42.9)	0(0)	1 (16.7)	1 (25.0)	
2: Moderate 3: Severe	2(28.6)	1 (14.3)	0(0)	1 (16.7) 0 (0)	1 (25.0) 0 (0)	
	1 (14.3)	0 (0)	0 (0)	0(0)	0(0)	
Inflammation						
Total	2 [2]	1.0 [1]	0 [0]	1[1]	1 [2]	0.13
1: Mild	2 (28.6)	5 (71.4)	5 (100)	4 (66.7)	3 (75.0)	0.28
2: Moderate 3: Severe	2 (28.6) 3 (42.9)	1 (14.3) 1 (14.3)	0 (0) 0 (0)	2 (33.3) 0 (0)	0 (0) 1 (25.0)	
	5 (42.9)	1 (14.3)	0(0)	0(0)	1 (23.0)	
Destruction	2.0 [2]	1.0 [2]	0 [0]	0 5 [1]	0.5 [1]	0.10
Total No Change	2.0 [2]	1.0[2]	0[0]	0.5 [1]	0.5 [1]	0.10
No Change Mild	2 (28.6) 1 (14.3)	3 (42.9) 2 (28.6)	5 (100) 0 (0)	3 (50.0) 3 (50.0)	2 (50.0) 2 (50.0)	0.11
Moderate	4 (57.1)	2 (28.6)	0 (0)	0 (0)	0 (0)	
	4 (37.1)	2 (20.0)	0(0)	0(0)	0 (0)	
Disorganization Total	2.0 [2]	1.0 [2]	0 [0]	0 [1]	05[1]	0.08
No Change	2.0 [2] 2 (28.6)	1.0 [2] 3 (42.9)	0 [0] 5 (100)	0 [1] 4 (66.7)	0.5 [1] 2 (50.0)	0.08
Mild	1 (14.3)	2 (28.6)	0 (0)	2 (33.3)	2 (50.0)	0.12
Moderate	4 (57.1)	2 (28.6)	0 (0)	0 (0)	0 (0)	
Crypt abscess						
Total	1.0 [2]	1.0 [1]	0 [0]	0.5 [1]	0.5 [1]	0.11
No Change	2 (28.6)	3 (42.9)	5 (100)	3 (50.0)	2 (50.0)	0.11
Mild	2 (28.6)	4 (57.1)	0 (0)	3 (50.0)	2 (50.0)	
Moderate	3 (42.9)	0 (0)	0 (0)	0 (0)	0 (0)	
		Bioch	emical results			
CRP (mg/dL)	2.11±1.26	3.51±3.56	1.87±0.20	1.98±0.20	2.32±0.63	0.53
SOD (units/mL)	5.49±2.57	5.50±2.24	3.84±2.15	3.45±1.66	2.70±1.41	0.13
WBC (×10 <sup>9</sup> /L)	6.21±3.46	8.10±6.71	8.22±3.15	11.50±5.39	10.18±5.60	0.43
HGB (g/dL)	12.36±1.10	12.54±1.54	12.16±2.28	13.17±1.80	11.50±1.28	0.61
PLT (×10 <sup>9</sup> /L)	874.86±335.16	397.20±443.22	600.20±621.02	796.33±430.63	628.25±476.44	0.37

Table 1 The effect of Melissa officinalis extracts on acetic acid-induced colitis in rats

\*The groups are described as: MO, received 300 mg/kg oral treatment with Melissa officinalis; MR, received 300 mg/kg rectal treatment with Melissa officinalis; AO received 100 mg/kg oral treatment with mesalamine; AR received 100 mg/kg rectal treatment with mesalamine; AA, the untreated animals with induced colitis

\*\*Fishers' exact or one-way analysis of variance/Kruskal-Wallis test

\*\*\*Scores are defined as 0, no macroscopic alterations; 1, just mucosal erythema; 2, mild mucosal edema, slight bleeding, or minor erosions; 3, moderate edema, slight bleeding ulcers, or erosions; 4, severe ulceration, edema, and tissue necrosis

CRP, C-reactive protein; HGB, hemoglobin; SOD, superoxide dismutase; WBC, white blood cells

127.5 mg of dried Melissa and 127.5 mg *Passiflora caerulea*, are used for the treatment of IBD, while also being effective

in the treatment of other gastrointestinal disorders, insomnia and anxiety [34,35]. On a macroscopic field and paraclinical evaluation, Melissa demonstrated similar properties to those of mesalamine, however, in our microscopic evaluation, Melissa, especially through the oral administration route, depicted poorer scores regarding ulceration, inflammation, destruction, disorganization, and crypt abscess, compared to the other groups. Protecting the colon structure from any pathologies caused by the inflammatory process, such as disorganization, adhesions, ulcerations, etc., is of the most important goals in the treatment of UC [26]. Therefore, based on the lack of studies regarding the effects of Melissa in the treatment of UC, and also our findings, further studies should be performed before the administration of these medications to human subjects.

Biochemical and laboratory changes were not significant in our study; however, we found lower levels of CRP and WBC, and higher SOD and HGB levels in the oral administration groups, compared to the control group. In the rectal administration groups, SOD and HGB levels improved. CRP improvement was only observed in the Melissa group, while WBC improvement was recorded in the mesalamine group. CRP, like other acute-phase reactive proteins, can have a negative effect on different phases of inflammation, which in our study was alleviated with the administration of Melissa. An increase in the SOD level improves colonic inflammation caused by UC [36], while our study showed that alleviation of bowel tract inflammation was achieved with the increase of SOD levels—a finding also supported by other studies on UC [37].

As a limitation of this study, some inflammatory mediators, such as colonic myeloperoxidase, colonic lipid peroxidation, colonic glutathione, and serum lactate dehydrogenase, which are sensitive markers for the inflammation of the bowel, were not evaluated [19]. Moreover, the net weight of the colonic specimen was not taken into account, and this is believed to be a sensitive and reliable marker for the extent and severity of the inflammatory response [38]. In addition, the stool consistency and degree of hematochezia were not documented; therefore, we are unable to add the disease activity index.

In conclusion, our study showed that *Melissa officinalis* has therapeutic effects against AA-induced UC in rats, particularly via topical administration. The advantages of these herbal remedies, given their lower reported adverse effects, should be taken into consideration. Therefore, further studies are needed to uncover the full potential and safety profile of the administration of this natural product as an alternative or complementary treatment in UC.

#### **Acknowledgments**

This study was supported by Shiraz University of medical sciences, Shiraz, Iran. The authors would also like to thank Dr. Vahedi for her help in the Animal House of Shiraz University of Medical Sciences, and all those who provided assistance during the study. We would also like to thank the assistance of Dr. Mohammad Hossein Khorraminejad-Shirazi in the preparations of the pathology slides.

#### Summary Box

#### What is already known:

- Ulcerative colitis (UC) is a form of chronic inflammatory bowel disease that affects the gastrointestinal tract and increases the chance of colorectal cancer
- Inflammation and oxidative activities within the gut play major roles in the pathogenesis of UC
- UC's current standard treatment has been shown to have several bothersome side-effect
- *Melissa officinalis*, an herb with anti-inflammatory, antioxidant and antibacterial effects, may be useful in diseases that are based on inflammatory and oxidative responses of the body, such as UC

#### What the new findings are:

- Administration of Melissa in an animal model showed significant results in reversing the damage to the intestinal mucosa due to acetic acid)
- In the group receiving oral administration of Melissa, 42.9% of animals demonstrated no macroscopic changes, while 14.3% demonstrated only mucosal erythema; there was no significant difference in the macroscopic features from the reference treatment group (mesalamine)
- Rectal administration of Melissa was superior to the reference group, although the difference was non-significant
- *Melissa officinalis* had therapeutic effects against acetic acid-induced UC in rats, particularly via topical administration

# References

- Jess T, Rungoe C, Peyrin-Biroulet L. Risk of colorectal cancer in patients with ulcerative colitis: a meta-analysis of population-based cohort studies. *Clin Gastroenterol Hepatol* 2012;10:639-645.
- 2. Wei SC, Sollano J, Hui YT, et al. Epidemiology, burden of disease, and unmet needs in the treatment of ulcerative colitis in Asia. *Expert Rev Gastroenterol Hepatol* 2021;**15**:275-289.
- Cosnes J, Gower-Rousseau C, Seksik P, Cortot A. Epidemiology and natural history of inflammatory bowel diseases. *Gastroenterology* 2011;140:1785-1794.
- Ng SC, Shi HY, Hamidi N, et al. Worldwide incidence and prevalence of inflammatory bowel disease in the 21<sup>st</sup> century: a systematic review of population-based studies. *Lancet* 2017;**390**:2769-2778.
- Crotty B, Jewell DP. Drug therapy of ulcerative colitis. Br J Clin Pharmacol 1992;34:189-198.
- Wen Z, Fiocchi C. Inflammatory bowel disease: autoimmune or immune-mediated pathogenesis? *Clin Dev Immunol* 2004;11:195-204.
- 7. Xu CT, Meng SY, Pan BR. Drug therapy for ulcerative colitis. *World J Gastroenterol* 2004;**10**:2311-2317.
- 8. Su C, Salzberg BA, Lewis JD, et al. Efficacy of anti-tumor necrosis factor therapy in patients with ulcerative colitis. *Am J Gastroenterol*

2002;97:2577-2584.

- Awaad AS, El-Meligy RM, Soliman GA. Natural products in treatment of ulcerative colitis and peptic ulcer. J Saudi Chem Soc 2013;17:101-124.
- Ye Q, Hu Z, Yang M, Qin K, Zhou Y. Effects and mechanisms of Chinese herbal medicine for ulcerative colitis: Protocol for a systematic review and meta-analysis. *Medicine (Baltimore)* 2020;99:e19768.
- 11. Zargari A. Medicinal plants. 6<sup>th</sup> Edition. Tehran University Publications, 1996.
- Mimica-Dukic N, Bozin B, Sokovic M, Simin N. Antimicrobial and antioxidant activities of *Melissa officinalis L. (Lamiaceae)* essential oil. J Agric Food Chem 2004;52:2485-2489.
- Kaufman T, Kalderon N, Ullmann Y, Berger J. Aloe vera gel hindered wound healing of experimental second-degree burns: a quantitative controlled study. J Burn Care Rehabil 1988;9:156-159.
- 14. Ashkani-Esfahani S, Ebrahimi A, Bahmani-Jahromi M, et al. The effect of *Melissa officinalis* extract on streptozotocin-induced diabetes in rats: a stereological study on pancreatic islets and betacells. *J Adv Med Biomed Res* 2021;**29**:34-40.
- 15. Saberi A, Abbasloo E, Sepehri G, et al. The effects of methanolic extract of *Melissa officinalis* on experimental gastric ulcers in rats. *Iran Red Crescent Med J* 2016;**18**:e24271.
- 16. Taiwo AE, Leite FB, Lucena GM, et al. Anxiolytic and antidepressant-like effects of *Melissa officinalis* (lemon balm) extract in rats: Influence of administration and gender. *Indian J Pharmacol* 2012;44:189-192.
- Charan J, Biswas T. How to calculate sample size for different study designs in medical research? *Indian J Psychol Med* 2013;35:121-126.
- Festing MF, Altman DG. Guidelines for the design and statistical analysis of experiments using laboratory animals. *ILAR J* 2002;43:244-258.
- Thippeswamy BS, Mahendran S, Biradar MI, et al. Protective effect of embelin against acetic acid induced ulcerative colitis in rats. *Eur J Pharmacol* 2011;654:100-105.
- 20. Fabia R, Willén R, Ar'Rajab A, Andersson R, Ahrén B, Bengmark S. Acetic acid-induced colitis in the rat: a reproducible experimental model for acute ulcerative colitis. *Eur Surg Res* 1992;24:211-225.
- 21. Tanideh N, Jamshidzadeh A, Ashraf M, Kuhi O, Mehrabani D. The healing effect of strawberry extract on acetic acid-induced ulcerative colitis in rat. *World Appl Sci J* 2014;**31**:281-288.
- 22. Tanideh N, Nematollahi SL, Hosseini SV, et al. The healing effect of *Hypericum perforatum* extract on acetic acid-induced ulcerative colitis in rat. *Ann Colorectal Res* 2014;**2**:e25188.
- 23. Misra HP, Fridovich I. The role of superoxide anion in the autoxidation of epinephrine and a simple assay for superoxide dismutase. *J Biol Chem* 1972;247:3170-3175.
- 24. Millar AD, Rampton DS, Chander CL, et al. Evaluating the antioxidant potential of new treatments for inflammatory bowel disease using a rat model of colitis. *Gut* 1996;**39**:407-415.

- 25. Murray A, Nguyen TM, Parker CE, Feagan BG, MacDonald JK. Oral 5-aminosalicylic acid for maintenance of remission in ulcerative colitis. *Cochrane Database Syst Rev* 2020;8:CD000544.
- Dignass A, Lindsay JO, Sturm A, et al. Second European evidencebased consensus on the diagnosis and management of ulcerative colitis part 2: current management. J Crohns Colitis 2012;6: 991-1030.
- Fan Y, Yi W, Huang H, Mei Z, Feng Z. Efficacy of herbal medicine (Gegen Qinlian Decoction) on ulcerative colitis: A systematic review of randomized controlled trials. *Medicine (Baltimore)* 2019;98:e18512.
- Huang C, Dong J, Jin X, et al. Intestinal anti-inflammatory effects of fuzi-ganjiang herb pair against DSS-induced ulcerative colitis in mice. *J Ethnopharmacol* 2020;**261**:112951.
- 29. Rezayat F, Hashempur MH, Tavahen H, Salmanroghani H, Emtiazy M. The efficacy of Ramak (a traditional herbal medicine preparation) for patients with ulcerative colitis: a pilot, randomized, triple-blinded, placebo-controlled clinical trial. *Eur J Integr Med* 2020;**39**:101209.
- 30. Sepehrimanesh M, Samimi N, Koohi-Hosseinabadi O, Mokhtari M, Amiri-Zadeh S, Farjam M. Effects of *Cupressus sempervirens* extract on the healing of acetic acid-induced ulcerative colitis in rat. J Coloproctol (Rio J) 2018;38:309-313.
- Ke F, Yadav PK, Ju LZ. Herbal medicine in the treatment of ulcerative colitis. Saudi J Gastroenterol 2012;18:3-10.
- 32. Świąder K, Startek K, Wijaya CH. The therapeutic properties of Lemon balm (*Melissa officinalis* L.): Reviewing novel findings and medical indications. J Appl Bot Food Qual 2019;92:327-335.
- 33. Dolatabadi F, Abdolghaffari AH, Farzaei MH, et al. The protective effect of *Melissa officinalis* L. in visceral hypersensitivity in rat using 2 models of acid-induced colitis and stress-induced irritable bowel syndrome: a possible role of nitric oxide pathway. *J Neurogastroenterol Motil* 2018;**24**:490-501.
- 34. Knop Laboratorios. Melipass. Available from: http://www. knoplabs.com/melipass/[Accessed 25 September 2023].
- Feliú-Hemmelmann K, Monsalve F, Rivera C. Melissa officinalis and *Passiflora caerulea* infusion as physiological stress decreaser. *Int J Clin Exp Med* 2013;6:444-451.
- 36. Seguí J, Gil F, Gironella M, et al. Down-regulation of endothelial adhesion molecules and leukocyte adhesion by treatment with superoxide dismutase is beneficial in chronic immune experimental colitis. *Inflamm Bowel Dis* 2005;**11**:872-882.
- Zhou YH, Yu JP, Liu YF, et al. Effects of Ginkgo biloba extract on inflammatory mediators (SOD, MDA, TNF-alpha, NF-kappaBp65, IL-6) in TNBS-induced colitis in rats. *Mediators Inflamm* 2006;**2006**:92642.
- Rachmilewitz D, Simon PL, Schwartz LW, Griswold DE, Fondacaro JD, Wasserman MA. Inflammatory mediators of experimental colitis in rats. *Gastroenterology* 1989;97:326-337.